The Nature of Bipolar Disorder

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The underlying pathophysiology of bipolar disorder is a continually evolving complexity of multilayer interacting and independent systems. The dearth of adequate preclinical or clinical models that incorporate the various features of the illness, i.e., acute and chronic, recurrent and episodic, and timecourse and treatment-related variables, has made the consistency and interpretation of data difficult. Newer technologies and the availability of structurally and mechanistically distinct pharmacologic agents have expanded opportunities for experimental study. In addition to the well-known neurotransmitter systems that are disrupted in mood disorders, critical guanine nucleotide–binding protein (G protein)–coupled signaling pathways are implicated in modulating mood state. Regulation of gene expression and identification of factors regulating neuroplasticity and neurotrophic events in the central nervous system in bipolar disorder are 2 of the more recent approaches contributing to clarification of the pathophysiology and potential treatment options.

C linical investigations of both the underlying pathophysiology and the treatment of bipolar disorder have expanded over the last 4 decades to encompass a diverse array of biochemical and neuroendocrine approaches and are now accessing basic fundamentals of gene regulation and expression. Early experimental and therapeutic strategies focused on dysregulations of the hypothalamic-pituitary-adrenal (HPA) axis and the monoaminergic systems as pivotal components of the manic and depressive states; however, this chronic affective disorder, which is characterized by unique cycling phenomena including progressive, episodic, and frequently severe mood disruptions, is now widely recognized to involve more complex biological interactions.

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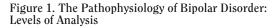
Space limitations necessitate our citing of review articles in many cases. A full reference list upon which this article was based is available upon request. (J Clin Psychiatry 2000;61[suppl 13]:42-57)

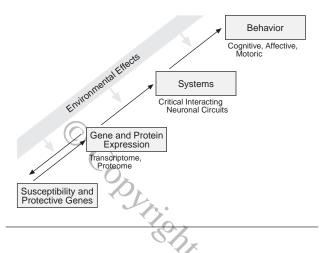
The underlying pathophysiology of bipolar disorder, partially elucidated through years of study with lithium and subsequently pursued through studies with the newer applications of antiepileptics such as carbamazepine, valproate, lamotrigine, and topiramate, should be understood through 4 descriptive physiologic levels: molecular, cellular, systemic, and behavioral (Figure 1).¹⁻⁴ These levels show complex interactions within components of each level as well as exert influences between levels. It is important to appreciate that the biological processes that are risk factors for some clinical manifestations of bipolar disorder, such as mood cycling, may be operating distinct from the factors manifesting as clinical symptoms of mania or depression per se.^{3,5,6} As genomic and molecular genetic studies of bipolar disorder progress, sophisticated experimental technology in analysis of gene expression and regulation is being applied to the neurobiology of bipolar disorder at the molecular levels; identification of specific gene sequences and associated factors that confer susceptibility or, equally importantly, exert a protective function in bipolar disorder will provide a starting point to follow the effects of the subsequent gene products through the different levels of physiologic outcome. The influences of these basic cellular and molecular mechanisms on critical neuronal circuitry and subsequent behavioral and clinical manifestations become dramatically complex as multiple signaling pathways in the limbic and limbicassociated regions of the brain contribute to the recurrent clinical affective symptomatology.¹

Beyond the complexity of the numerous independent and associated biological pathways that are apparent in various aspects of the manic and depressive phases of the illness, concurrent physiologic feedback systems are undoubtedly activated in an attempt to compensate for the

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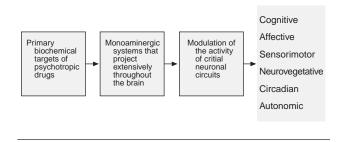
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extreme fluctuations in normal pathways and to restore normal homeostasis. The array of molecular and cellular abnormalities and the compensatory mechanisms for these abnormalities may also develop longitudinally and will vary in conjunction with the stage and progression of illness. As a result of potential heterogeneity in these dysfunctional and response systems, individual patients may present with a unique collection of clinical symptoms consisting not only of mood disturbances but also of endocrine, circadian, and drug treatment-related patterns (Figure 2). Furthermore, the acute versus chronic nature of bipolar disorder, with frequently unpredictable recurrence and cycling between the phases, adds additional layers of difficulty in identifying consistent or comparable model systems or patient groups for study. However, while precise knowledge of the etiology and pathophysiology of this disorder that affects millions is still limited, it has become increasingly clear that elucidation of the signaling pathways affected in bipolar disorder will greatly facilitate the development of novel, improved therapeutics. Furthermore, advancing our knowledge of the underlying neurobiology of the illness and the neuronal circuits mediating disease and treatment manifestations will undoubtedly facilitate a much better understanding about the precise manner in which the susceptibility and protective genes interact with each other as well as with environmental factors to produce one of the most devastating and at the same time intriguing of all neuropsychiatric illnesses-bipolar disorder.

While the efficacy of lithium as a mood stabilizer has generated a large body of clinical and experimental findings in bipolar disorder, it has become increasingly appreciated that a substantial minority of patients fail to respond, or respond only partially, to lithium therapy. Several reports indicate that as many as 20% to 40% of patients either do not attain effective therapeutic remission with lithium or continue to incur significant morbidity.

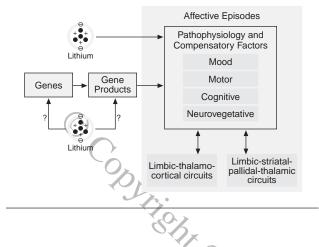


Thus, a critical task becomes the development of newer therapeutic agents that will have a quicker, more potent, and more specific mode(s) of action with fewer side effects, thereby enhancing compliance. Although the study of lithium has provided significant mechanistic information on physiologic and biochemical factors and systems associated with mood perturbation, the discovery and application of newer agents including carbamazepine, valproate, lamotrigine, topiramate, risperidone, clozapine, and olanzapine open additional avenues for clarifying the fundamentals of mood disorders at the molecular and cellular level. A review of the progress in elucidating the biochemical and genetic targets that may protect or predispose an individual to bipolar disorder also serves to reinforce the complexity of the multiple independent and overlapping cascades of neurotransmitter and signal transduction systems that may be responsible for the clinical symptoms of depression and mania. In this article, we have attempted to highlight some of the most relevant data over the years pertaining to the complex neurobiology involved in the pathophysiology of bipolar disorder. For the sake of brevity, we sometimes reference review articles.

NEUROTRANSMITTERS

The original "monoamine hypothesis" describing the relationship between excesses or deficiencies in monoamine neurotransmitter concentration in the central nervous system (CNS) and affective disorders (depression and mania) has been revised to include the role of the cellular receptors for the biogenic amines in these 2 states.^{7–11} Not only have recent pharmacologic agents that were developed in the burgeoning era of the monoamine hypothesis been applied with definitive clinical effects on single episodes of mania or depression, but they have also provided research tools to further probe the biology of bipolar disorder. Additionally, the potential ability of antidepressants to trigger manic episodes and promote rapid cycling has led to insights into the pathophysiology of the illness.¹²⁻¹⁴ Since the pathophysiologic pattern of bipolar disorder most likely involves dysfunction in the limbic

Figure 3. The Pathophysiology of Bipolar Disorder: Clues From the Molecular and Cellular Actions of Lithium



and limbic-related systems, which are extensively regulated by the biogenic amine neurotransmitter systems, these systems are prominently placed to mediate the complex behavioral manifestations of the disease and also serve as therapeutic targets (Figure 3). While these concepts seem straightforward, the multifunctional nature of neurotransmitter systems, the plethora of available neurotransmitter and neuropeptide candidates, and the surfeit of biochemical data complicate assignment of definitive meaning to the observed abnormalities (Table 1).

Noradrenergic System

The catecholamine hypothesis of mood disorders has been extensively investigated^{7,8,15} but has proved difficult to study experimentally owing, at least in part, to the formidable methodological difficulties in assessing CNS noradrenergic function in humans, as noted above. Overall, however, the data in toto do suggest an abnormality of the noradrenergic system in bipolar disorder.^{5,15,16} Studies of plasma norepinephrine suggest that, on average, bipolar depressed patients appear to have reduced-to-normal resting output of norepinephrine, with a highly exaggerated noradrenergic system response to standing. Plasma 3methoxy-4-hydroxyphenylglycol (MHPG) levels also tend to be lower in bipolar than in unipolar depressed patients and are higher in bipolar patients when they are manic than when they were depressed.^{5,13}

For some years, measures of norepinephrine and its metabolites in cerebrospinal fluid (CSF) were thought to directly reflect the activity of brain noradrenergic systems. This assumption is, however, problematic, since high correlations have been found between indices of norepinephrine function in plasma and CSF.¹⁷ Certain other methodological problems are unique to CSF measures. Standards for obtaining spinal fluid, such as elapsed time between needle insertion and sample collection, have not been established.^{5,16} Therefore, subjects may not have the same

Table 1. The Pathophysiology of Mood Disorders: A Surfeit of Biochemical Findings^a Noradrenergic System Urinary MHPG levels Plasma norepinephrine levels CSF norepinephrine, MHPG levels Peripheral and central α_2 -adrenergic receptors, β-adrenergic receptors Neuroendocrine challenges AMPT effects Dopaminergic System CSF HVA levels Electroconvulsive therapy effects AMPT challenges Psychomotor retardation and activation Cholinergic System Serotonergic System CSF 5-HIAA levels Neuroendocrine challenges Peripheral and central 5-HT₂ receptors [³H]imipramine, [³H]paroxetine binding PET findings on decreased 5-HT_{1A} binding Tryptophan depletion effects Corticotropin-Releasing Factor (CRF) HPA axis hyperactivity CRF in CSF Pituitary and adrenal enlargement Postmortem brain CRF receptors Circadian Rhythms Regional Fluorodeoxyglucose Uptake ^aAbbreviations: AMPT = α -methylparatyrosine, CSF = cerebrospinal fluid, 5-HIAA = 5-hydroxyindoleacetic acid, HPA = hypothalamicpituitary-adrenal, 5-HT = 5-hydroxytryptamine, HVA = homovanillic acid, MHPG = 3-methoxy-4-hydroxyphenylglycol, PET = positron emission tomography.

degree of accommodation to the stress of the needle stick. Moreover, sampling at a single point in time may not reflect the biochemical process of depression or mania but rather a state-dependent fluctuation from a recent external or internal stress.^{5,16}

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Since the catecholamine hypothesis of mood disorders was proposed, attempts to characterize output of the noradrenergic system in depressed patients have focused on cumulative measurements of MHPG in urine more than any other single parameter. Detailed reviews of the literature differ in their conclusions, but overall, it appears that urinary MHPG levels are lower in bipolar depressed patients, and—similar to the plasma studies—longitudinal studies show that MHPG excretion is higher in the manic compared with the depressed state.^{5,15,16,18}

Cerebrospinal fluid norepinephrine is reported to be higher in mania than in depression, and CSF MHPG appears to be lower in bipolar depression than in unipolar depression.¹⁶ Additionally, CSF MHPG has also been reported to be elevated in manic patients compared with controls and to be reduced with lithium treatment, even when treatment was unsuccessful. Taken together, the CSF studies of norepinephrine and its metabolite MHPG suggest that norepinephrine output is higher in mania than in depression and that there may be relatively higher values in unipolar versus bipolar depression. Over the years, a number of studies have also attempted to infer alterations in central adrenergic receptor function in mood disorders using neuroendocrine challenge paradigms as well as studies of adrenergic receptors on platelets and lymphocytes.^{13,15} The caveats necessary when generating elaborate hypotheses about CNS adrenergic receptor dysfunction solely on the basis of studies of these tissues have already been articulated. In the aggregate, these studies *do* tend to suggest altered sensitivity of α_2 and β_2 -adrenergic receptors in mood disorders. However, at present, a parsimonious interpretation is that they represent the sequelae, not the cause, of other, more primary perturbations.

Serotonergic System

As with the noradrenergic system, interest in the role of the serotonergic system in mood disorders derived from a long-standing tradition of research into the role of this indolamine in the mechanisms of action of effective treatments. A consistent body of data from CSF studies, neuroendocrine challenge studies, serotonin receptor and reuptake site binding studies, pharmacologic studies, and most recently, brain imaging studies support a role for alterations of serotonergic neurotransmission in depression.^{5,19,20} The role of the serotonergic system in bipolar disorder, however, is less clear. Overall, investigators have reported reduced levels of 5-hydroxyindoleacetic acid (5-HIAA) in a subgroup of patients, especially those with impulsivity, aggression, and suicide attempts.^{5,19}

A variety of studies using different methodological and technical approaches have devoted attention to the status of serotonin receptors in depression: decreased radioligand binding to the 5-hydroxytryptamine (5-HT) receptor in the platelets and midbrain of unipolar patients and increases in the density of serotonin-2 (5-HT₂) receptors in platelet and postmortem brain studies in depression.^{19,20} Most recently, an intriguing preliminary positron emission tomography study²¹ reported decreases in 5-HT_{1A} binding potential in several areas of brain in depressed patients, in particular in bipolar depressives and in unipolar patients who have bipolar relatives. Although the small sample size precludes a definitive interpretation of these findings, the use of similar receptor-specific and postreceptorspecific radioligands in future studies should greatly enhance our understanding of the pathophysiology of mood disorders. Distribution of the multiple, well-characterized 5-HT receptor subtypes is known to vary throughout the peripheral and central nervous system, and manipulation of the activity of these receptors in different disorders may have different primary and downstream outcomes. However, reasonable elucidation of these pathways may augment the basic understanding of the pathophysiology of mood disorders.¹¹ An emerging number of genetic studies are hinting at an association between polymorphisms in the serotonin transporter and tryptophan hydroxylase

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genes and features of mood disorders; however, these findings require extensive further validation.

Neurotransmitter depletion models, specifically in this case tryptophan depletion to lower serotonin levels, permit a more direct strategy to clarify the involvement of serotonergic systems in mood disorders. Tryptophan depletion results in reversal of the response to selective serotonin reuptake inhibitor (SSRI) (but not noradrenergic) antidepressant medications and recurrence of depression; however, depletion in healthy patients without evidence of mental illness and in nonmedicated patients with depression did not cause or intensify depression.²² These studies again substantiate the underlying complexity of neurobiological systems inherent not only in depression but, by analogy, in bipolar illness. Two small studies^{23,24} have investigated the effect of tryptophan depletion in recently manic, lithium-treated euthymic patients, with conflicting results.

Dopaminergic System

Several lines of evidence point to a role for dopamine in mood disorders. A relevant preclinical model derives from the crucial role of mesoaccumbens dopamine in the neural circuitry of reward and/or incentive motivational behavior.^{25,26} Loss of motivation is one of the central features of depression, and indeed, anhedonia is one of the defining characteristics of melancholia; thus, a deficiency of dopamine systems stands out as a prime candidate for involvement in the pathophysiology of depression.^{25,26} The strongest direct finding from clinical studies implicating dopamine in depression is reduced homovanillic acid (HVA, the major dopamine metabolite) in the CSF; indeed, this is one of the most consistent biochemical findings in depression.^{5,16} There is also evidence for a decreased rate of CSF HVA accumulation in subgroups of depressed patients, including those with marked psychomotor retardation versus those with agitation.²⁶ Furthermore, depression occurs in up to 40% of patients with idiopathic Parkinson's disease and may precede motor symptoms.

The pharmacologic bridge also supports the notion that manipulation of the dopaminergic system is capable of modulating the illness. Thus, dopamine agonists appear to be effective antidepressants in some bipolar patients and are also able to precipitate mania in some bipolar patients.^{5,13} Analyzing the effects of pharmacologic manipulations of the dopaminergic and noradrenergic systems, Goodwin and Sack²⁷ postulated that dopamine abnormalities are primarily involved in the hyperactivity and psychosis associated with the more severe stages of mania, whereas norepinephrine might be implicated in the euphoria-grandiosity more characteristic of hypomania. They heuristically envisioned a cascade in which the initial noradrenergic dysregulation of hypomania gives way to the hyperdopaminergic state of mania. Linkage of biochemical evidence and behavioral observation during measurement of psychomotor function and its relationship to catecholamine function in bipolar patients as evaluated by monitoring concentrations of catecholamines and their metabolites in the CSF and urine has shown that increased arousal and prominent psychomotor abnormalities are associated with the catecholamine system.²⁸ Investigators have utilized a catecholamine depletion strategy (most recently, via use of the tyrosine hydroxylase inhibitor α -methyl-ptyrosine [AMPT]) to elucidate the putative roles of dopamine and norepinephrine in mood disorders. In addition to inducing a recurrence of symptoms in depressed patients being treated with "noradrenergic antidepressants," AMPT may induce depressive symptoms in some euthymic, medication-free patients.^{15,29} Most recently, investigators have examined the effects of AMPT administration on lithium-treated, euthymic bipolar disorder patients.³⁰ Intriguingly, they observed no mood-lowering effects of AMPT but observed a rebound hypomania in a significant percentage of the patients. Although preliminary, these results are compatible with a dysregulated signaling system wherein the compensatory adaptation to catecholamine depletion results in an "overshoot" due to impaired homeostatic mechanisms.

Cholinergic System

Much of the evidence supporting the involvement of the cholinergic system in some areas of mood disorders comes from neurochemical, behavioral, and physiologic studies in response to pharmacologic manipulations in investigations done in the early 1970s showing that the relative inferiority of noradrenergic compared with cholinergic tone was associated with depression, while conversely, the opposite was associated with mania.9,10 Additional support is found from a study of the central cholinesterase inhibitor physostigmine (administered intravenously) in which transient modulation of symptoms in manic patients and induction of depression in euthymic bipolar patients stabilized with lithium were observed.¹⁰ Further extensive studies on the effects of lithium to perhaps potentiate brain cholinergic systems have added to the evidence.¹ However, the therapeutic responses observed with antidepressant and antimanic pharmacologic agents do not correlate with their respective efficacy on the cholinergic system.

Hypothalamic-Pituitary-Adrenal Axis

A large body of data has documented abnormalities of the HPA axis in mood disorders.³¹ Numerous reports document HPA axis hyperactivity in drug-free depressed and bipolar depressed patients. With respect to bipolar disorder, increased HPA activity has been associated with mixed-manic states, depression, and, less consistently, classic manic episodes.^{5,20} Depression is associated with perturbation in HPA axis function, as observed through increased baseline HPA axis function (higher corticotropin-releasing factor [CRF] in CSF, increased plasma adrenocorticotropic hormone [ACTH] and cortisol concentrations) as well as failure to suppress plasma ACTH and cortisol during the dexamethasone suppression test (DST).¹⁹ In patients diagnosed with severe depression, effective pharmacotherapy of the depression is linked to normalization of the hyperactive HPA axis and resultant mood elevation.³² The DST has also been studied in patients with bipolar disorder; approximately 25% to 60% of depressed bipolar patients are reported to have abnormal DST results, with some (but not all) studies reporting a normalization of the results during the hypomanic or manic phases. The recent development of CRF antagonists for the treatment of depression will allow more direct clinical and experimental investigations into the role of CRF hypersecretion in the pathophysiology of depression.

The potential hyperactivation of the HPA axis in mood disorders has been revisited in recent years, in large part owing to the growing recognition of the brain atrophy that may be present in many patients (discussed later). It remains to be fully elucidated to what extent these findings represent the sequelae of the biochemical changes (for example, in glucocorticoid levels) accompanying repeated affective episodes per se. The suggestion that changes in glucocorticoid levels affect brain atrophy receives support from the observation that chronic stress or glucocorticoid administration has been demonstrated to produce atrophy and death of vulnerable hippocampal neurons in rodents and primates. Furthermore, magnetic resonance imaging (MRI) studies have also revealed reduced hippocampal volumes in patients with Cushing disease and posttraumatic stress disorder (other conditions associated with hypercortisolemia). Indeed, one of the most consistent effects of stress on cellular morphology is atrophy of hippocampal neurons.^{33,34} This atrophy is observed in the CA3 pyramidal neurons, but not in other hippocampal cell groups (i.e., CA1 pyramidal and dentate gyrus granule neurons). Atrophy of CA3 pyramidal neurons also occurs upon exposure to high levels of glucocorticoids, suggesting that activation of the HPA axis very likely plays a major role in mediating the stress-induced atrophy. In addition to neuronal atrophy, longer-term exposure to stress (i.e., for several months) can result in true loss of hippocampal neurons in the CA3 pyramidal cell layer.³³ Furthermore, it is possible that recurrent mood disorders may lower the threshold for cell death/atrophy in response to a variety of other physiologic (e.g., normal aging) and pathologic (e.g., ischemia) events and thereby contribute to a variety of deleterious health-related effects.

SIGNAL TRANSDUCTION NETWORKS

Signal transduction networks, composed of membranebound and intracellular complexes of multiple protein, enzyme, and small molecule components, transmit extracellular signals to intracellular processing systems to direct the eventual cellular response to stimulation or inhibition.³⁵⁻³⁷ These signaling networks mediate communications over a variety of time scales; thus, in addition to mediating instantaneous responses to receptor activation, signaling systems also participate in long-term neuroplastic events and regulate cellular memory. Cells that have experienced different stimuli may express a unique repertoire of signaling molecules and transcription factors, allowing for different responses to the identical stimuli. Signal transduction networks can amplify or redirect the initial stimuli and regulate both message propagation and feedback loops and thereby serve major integrative functions to form both intracellular and intercellular circuits. Given the important roles of signal transduction networks in regulating normal physiology as well as identifying signaling abnormalities in a number of disease states, their involvement in the pathophysiology and treatment of bipolar disorder would not be surprising. Pertinent for the present discussion is the observation that a variety of diseases arising from abnormalities in guanine nucleotidebinding protein (G protein)-coupled signal transduction pathways manifest relatively circumscribed symptomatology,^{38,39} despite the widespread, often ubiquitous expression of the affected signaling proteins. Moreover, recent research has clearly identified signaling pathways as therapeutically relevant targets for the most effective pharmacologic treatments.^{40–44} Indeed, the molecular and cellular targets underlying lithium's ability to stabilize an underlying dysregulation of limbic and limbic-associated functions strongly suggest that abnormalities in signaling pathways may also play a critical role in the pathophysiology of bipolar disorder.45 We now turn to a discussion of the direct and indirect evidence supporting a role for abnormalities in signaling pathways in the pathophysiology of bipolar disorder.

The Cyclic Adenosine Monophosphate/ Protein Kinase A Signaling Pathway

The most commonly utilized strategy in patients with mood disorders has been to focus on receptor function in readily accessible blood elements, and much clinical research has focused on the activity of the cyclic adenosine monophosphate (cAMP) generating system in mood disorders. Overall, the preponderance of the evidence suggests altered receptor and/or postreceptor sensitivity of the cAMP generating system in mood disorders in the absence of consistent alterations in the number of receptors themselves.^{46,47} A recent study^{48,49} found higher levels of cAMPstimulated phosphorylation of a ~22 kDa protein in platelets obtained from 10 treated euthymic bipolar disorder patients compared with healthy subjects; by contrast, no significant difference in basal phosphorylation was found between the groups. Follow-up studies identified the ~22 kDa protein as Rap1 and once again found higher cAMPstimulated phosphorylation in the bipolar disorder patients.^{48,49} Rahman and associates⁵⁰ have undertaken the most thorough series of studies investigating the cAMP/ protein kinase A (PKA) system in postmortem human brain in bipolar disorder. They found that the levels of PKA regulatory subunits (as assessed by [³H]cAMP binding) were significantly lower in cytosolic fractions of bipolar disorder frontal, temporal, occipital, and parietal cortex; cerebellum; and thalamus compared with matched controls. Furthermore, preliminary findings show that the reduction of regulatory subunits of PKA in the cytosolic fractions of bipolar disorder temporal cortex is accompanied by a higher basal kinase activity and significantly lower apparent activation constant for cAMP in the cytosolic fractions of bipolar disorder temporal cortex.⁵¹ These observed changes in PKA provide additional important evidence for dysregulation in the $G\alpha_s$ -mediated cAMP cascade in bipolar disorder.

Lithium and the cAMP Generating System

Lithium has been demonstrated to exert complex effects on the activity of adenylyl cyclase (AC), with the preponderance of the data demonstrating an elevation of basal AC activity while attenuating a variety of receptormediated responses.^{46,52–54} Lithium in vitro inhibits the stimulation of AC by guanyl imidodiphosphate, or Gpp(NH)p (a poorly hydrolyzable analogue of guanosine triphosphate [GTP]), and calcium-calmodulin, both of which can be overcome by Mg²⁺.55-57 Lithium also competes with Mg²⁺ for membrane-binding sites, and the inhibition of solubilized catalytic unit of AC by lithium can also be overcome by Mg²⁺. These findings suggest that lithium's inhibition of AC in vitro may be due to competition with Mg²⁺ on a site on the catalytic unit of AC.^{55,57} However, the inhibitory effects of chronic lithium treatment on rat brain AC are not reversed by Mg²⁺, and these inhibitory effects still persist after washing of the membranes but are reversed by increasing concentrations of GTP.⁵⁶ These results suggest that the physiologically relevant effects of lithium (i.e., those seen with chronic drug administration and not reversed immediately with drug discontinuation) may be exerted at the level of signaltransducing G proteins at a GTP-responsive step (discussed later). These 2 distinct actions of lithium on the AC system may explain the differing results obtained by investigators using rat membrane preparations or slice preparations.^{43,44} These results have led to an investigation of the effects of lithium on the AC system in vivo, using microdialysis. These studies found that chronic lithium treatment produced a significant increase in basal and postreceptor-stimulated (cholera toxin or forskolin [FSK]) AC activity, while attenuating the β -adrenergic-mediated effect.43,44,58 Interestingly, chronic lithium treatment resulted in an almost absent cAMP response to pertussis toxin, suggesting a lithium-induced attenuation of G_i function. It should be noted, however, that chronic lithium has also been found to increase not only cAMP levels,⁵⁹ but

also the levels of AC Type I and Type II mRNA and protein levels in frontal cortex,^{60,61} suggesting that the complex effects of lithium on the system may represent the net effects of direct inhibition of AC, up-regulation of AC subtypes, and effects on the stimulatory and inhibitory G proteins. Most recently, the effects of lithium on the phosphorylation and activity of CREB (cAMP response element binding protein) have been examined in rodent brain and in cultured human neuroblastoma cells, with somewhat conflicting results.⁶²⁻⁶⁴

Another series of studies has examined the effects of lithium on AC in humans. In a longitudinal study of healthy volunteers, 2 weeks of lithium administration was found to significantly increase platelet basal and postreceptor-stimulated AC activity,⁶⁵ effects that are strikingly similar to those observed in rodent brain. Consistent with a lithium-induced increase in basal cAMP and AC levels, a more recent study⁴⁸ found that platelets obtained from lithium-treated euthymic bipolar patients enhanced basal and cAMP-stimulated phosphorylation into Rap1 (a PKA substrate) as well as into a 38 kDa phosphoprotein. Interestingly, these investigators did not find similar effects of lithium in healthy subjects.

Carbamazepine and the AC System

Considerable data suggest that carbamazepine (an atypical anticonvulsant) may be an effective alternative or adjunctive treatment to lithium, both for acute manic episodes as well as for long-term prophylaxis in bipolar disorder. Despite the widespread clinical use of carbamazepine, the cellular mechanism(s) underlying both its anticonvulsant and mood-stabilizing effects have not been fully elucidated.^{66,67} In contrast to the effects observed with lithium and valproate described above, it has been demonstrated that carbamazepine has very modest effects on the protein kinase C (PKC) signaling pathway and G proteins (H.K.M., G. Chen, M.D., Ph.D., unpublished observations, October 1997). Carbamazepine has, however, been demonstrated to have many effects on the cAMP signaling pathway.

Carbamazepine has been demonstrated to decrease not only basal, but also norepinephrine-, adenosine-, veratridine-, and ouabain-stimulated cAMP levels in rodent brain.⁶⁸⁻⁷⁴ Recent studies have also demonstrated that carbamazepine inhibits forskolin-induced c-*fos* gene expression in cultured pheochromocytoma (PC12) cells.⁷⁵ Thus, overall, considerable evidence indicates that carbamazepine inhibits cAMP formation. Studies have recently been undertaken to investigate the possible mechanisms by which carbamazepine inhibits the cAMP generating system. It was found that carbamazepine, at therapeutically relevant concentrations, inhibited both basal AC and FSKstimulated cAMP accumulation in C6 glioma cells.⁷⁶ To further characterize the site at which carbamazepine exerts its inhibitory effects, ACs were purified from rat cerebral

cortex using a FSK affinity purification column. Carbamazepine inhibited both basal and FSK-stimulated activity of purified AC.⁷⁶ Taken together, the data suggest that carbamazepine inhibits cAMP production by acting directly on AC and/or through factor(s) that are tightly associated with, and copurify with, AC. It is now well established that the antimanic effects of carbamazepine show a lag period of onset. In this context, it is noteworthy that Divish and colleagues⁷⁵ have demonstrated that carbamazepine attenuates FSK-induced c-fos (an immediate early gene) expression in PC12 cells. Since c-fos and other immediate-early genes are known to be involved in mediating a number of long-term neuronal responses,⁷⁷ these effects might be postulated to play a role in the delayed therapeutic effect of carbamazepine. Consistent with these observations, it was found that in C6 glioma cells, FSK enhances phosphorylation of CREB and that carbamazepine significantly inhibited FSK-induced phosphorylation of CREB. These results complement the results of Divish and associates⁷⁵ described above and suggest that the inhibitory effects of carbamazepine on AC have the potential to bring about long-term changes in gene expression and thereby regulate cellular function.

The Phosphoinositide/PKC Signaling Pathway

Peripheral cell studies and postmortem brain studies have generally revealed modest abnormalities in the phosphoinositide/PKC signaling system in bipolar disorder. Thus, one study⁷⁸ measured membrane phospholipids in platelets of 7 medication-free patients in the manic phase of bipolar disorder and 7 healthy comparison subjects. The relative percentage of platelet membrane phosphatidylinositol 4,5-bisphosphate (PIP₂) was significantly higher in the manic patients than in the comparison subjects. More recently in a single case study,⁷⁹ the same laboratory studied PIP, membrane values in a bipolar disorder patient in different mood states. They found that the relative percentage of PIP₂ in the platelet membranes increased with cycling from the euthymic into the manic state. After lithium treatment, PIP, decreased and was similar to the euthymic state. Consistent with these findings, van Calker and associates⁸⁰ have found increased sensitivity to agonist stimulation of the Ca²⁺ response in neutrophils of manic depressive patients, effects that were normalized by lithium treatment. Investigators have also attempted to determine if the altered phosphoinositide signaling found in peripheral cells from bipolar disorder patients is present in the CNS. In this context, Mathews and associates⁸¹ found increased $G\alpha_{\alpha/11}$ immunoreactivity in postmortem occipital cortex from patients with bipolar disorder. However, these elevated levels of $G\alpha_{a/11}$ were accompanied by reduced agonist-induced phosphoinositide turnover,⁸² although the potential effects of long-term lithium treatment remain to be fully delineated. To date, few studies have directly examined PKC in bipolar disorder.

Friedman and associates⁸³ investigated PKC activity and translocation in response to serotonin in platelets obtained from bipolar disorder subjects before and during lithium treatment. They found that the ratios of membrane-bound to cytosolic PKC activities were elevated in platelets obtained from the manic subjects, compared with patients with schizophrenia and healthy controls. In addition, serotonin-stimulated platelet PKC translocation was found to be enhanced in manic subjects. Moreover, lithium treatment for up to 2 weeks resulted in a reduction in cytosolic and membrane-associated PKC activities and in an attenuated PKC translocation in response to serotonin. These preliminary results suggest that alteration in platelet PKC is associated with the manic phase of bipolar illness. More recently, the same laboratory has measured PKC isozyme levels, activity, and translocation in postmortem brain tissue from bipolar disorder patients; they have found increased PKC activity and translocation as well as elevated levels of cytosolic α - and membrane-associated PKC γ and PKC ζ isozymes in brains obtained at postmortem from bipolar disorder patients compared with controls.^{84,85} These results may be specific to bipolar disorder, since another laboratory has recently found significantly decreased phorbol dibutyrute ([3H]PDBU) binding sites in both membranous and cytosolic postmortem brain samples. (Brodmann areas 8 and 9) obtained from teenage suicide victims compared with control subjects.⁸⁶

Lithium and PKC

The "inositol depletion hypothesis" posited that lithium, as an uncompetitive inhibitor of inositol-1-phosphatase (IMPase), produced its therapeutic effects via a depletion of neuronal myo-inositol levels. Although this hypothesis has been of great heuristic value, numerous studies have examined the effects of lithium on receptor-mediated phosphoinositide responses, and although some report a reduction in agonist-stimulated PIP₂ hydrolysis in rat brain slices following acute or chronic lithium administration, these findings have often been small, inconsistent, and subject to numerous methodological differences.^{42,87} Most recently, a proton magnetic resonance spectroscopy (MRS) study⁸⁸ has demonstrated that lithium-induced myoinositol reductions are observed in the frontal cortex of bipolar disorder patients after only 5 days of lithium administration, at a time when the patients' clinical state is completely unchanged. Consequently, these and other studies suggest that while inhibition of IMPase may represent an initiating lithium effect, reducing myo-inositol levels per se is not associated with therapeutic response. This finding has led to the working hypothesis that some of the initial actions of lithium may occur with a relative reduction of myo-inositol; this reduction of myo-inositol initiates a cascade of secondary changes in the PKC signaling pathway and gene expression in the CNS, effects that are ultimately responsible for lithium's therapeutic efficacy.

Table 2. Direct and Indirect Evidence Supporting a Role for Protein Kinase C (PKC) in the Pathophysiology and Treatment of Bipolar Disorder^a

- Kindling produces dramatic increases in membrane-associated PKC in hippocampus and amygdala
- Amphetamine produces increases in PKC activity and GAP-43 phosphorylation (implicated in neurotransmitter release)
- PKC inhibitors block the biochemical and behavioral responses to amphetamine and cocaine and also block cocaine-induced sensitization
- Dexamethasone administration increases PKC activity and the levels of PKC α and PKC ϵ in rat frontal cortex and hippocampus
- Membrane/cytosol PKC partitioning in platelets is increased in manic subjects; normalizes with lithium treatment
- Bipolar patients' brains show increased PKC activity and translocation compared with controls
- Lithium and valproate regulate PKC activity, PKC $\alpha,$ PKC $\epsilon,$ and MARCKS
- Preliminary data suggest that PKC inhibitors may have efficacy in the treatment of acute mania
- Typical and atypical antipsychotics regulate the levels of PKC α and ϵ in areas of rat brain

^a Abbreviations: GAP = growth cone–associated protein,	
MARCKS = myristoylated alanine-rich C kinase substrate.	

Indeed, evidence accumulating from various laboratories has clearly demonstrated that lithium, at therapeutically relevant concentrations, exerts major effects on the PKC signaling cascade.^{40-42,85,89} PKC is highly enriched in brain and plays a major role in regulating presynaptic and postsynaptic aspects of neurotransmission. PKC is one of the major intracellular mediators of signals generated upon external stimulation of cells via a variety of neurotransmitter receptor subtypes, which induce the hydrolysis of membrane phospholipids. PKC is now known to exist as a family of closely related subspecies, has a heterogeneous distribution in brain (with particularly high levels in presynaptic nerve terminals), and plays a major role in the regulation of neuronal excitability, neurotransmitter release, and long-term alterations in gene expression and plasticity (Table 2).

The preponderance of the currently available data suggests that acute lithium may activate PKC, whereas chronic lithium results in an attenuation of phorbol estermediated responses accompanied by a down-regulation of PKC isozymes in the brain. Using quantitative autoradiographic techniques, it was demonstrated that chronic (5-week) lithium administration results in a significant decrease in membrane-associated PKC in several hippocampal structures, most notably the subiculum and the CA1 region, in the absence of any significant changes in the various other cortical and subcortical structures examined.90,91 Furthermore, immunoblotting using monoclonal anti-PKC antibodies has revealed isozyme-specific decreases in PKC α and PKC ϵ (which have been particularly implicated in facilitating neurotransmitter release), in the absence of significant alterations in PKC β , PKC γ , PKC δ , or PKC ζ . It is also noteworthy that exposure of immortalized hippocampal cells,³⁷ neuroblastoma cells, or

PC12 cells92 to lithium (1.0 mM) in vitro produces isozyme-selective decreases in PKC α and (in the case of PC12 cells) PKC ε. In the absence of suitable animal models for the major psychiatric disorders, a major problem inherent in neuropharmacologic research is the difficulty in precisely ascribing therapeutic relevance to any observed biochemical finding. One approach that has been utilized is the identification of common biochemical targets that are modified by drugs belonging to the same therapeutic class (e.g., antimanic agents) but possessing distinct chemical structures (i.e., lithium, valproate, and olanzapine, the only 3 drugs approved by the Food and Drug Administration for the treatment of bipolar disorder) when administered in a "therapeutically relevant" paradigm. Although they most likely do not work by precisely the same mechanisms, identifying the biochemical targets that are regulated in concert by these 2 chemically distinct agents may provide important clues about molecular mechanisms underlying mood stabilization in the brain.

In view of the significant effects of lithium on PKC, outlined above, the effects of valproate on various aspects of PKC functioning have also been investigated. It has been found that the structurally highly dissimilar agent valproate produces effects on the PKC signaling pathway that are strikingly similar to lithium's effects.^{42,93} Interestingly, chronic lithium and valproate appear to regulate PKC isozymes by distinct mechanisms, with valproate's effects appearing to be largely independent of myoinositol. This biochemical observation is consistent with the clinical observations that some patients show preferential response to one of the agents and that one often observes additive therapeutic effects in patients when the 2 agents are coadministered. In view of the pivotal role of the PKC signaling pathway in the regulation of neuronal excitability, neurotransmitter release, and long-term synaptic events, we postulated that the attenuation of PKC activity may play a role in the antimanic effects of lithium and valproate. In a pilot study,94 it was found that tamoxifen (a nonsteroidal antiestrogen known to be a PKC inhibitor at higher concentrations) may, indeed, possess antimanic efficacy. While awaiting further replication, these data further support the potential role of the PKC pathway in mediating the pathophysiology of bipolar disorder.

A major strategy that has been utilized to investigate the downstream consequences of lithium-induced alteration in PKC isozymes is the examination of the effects of chronic lithium on endogenous PKC substrates in brain. The most prominent substrate for PKC in brain is an acidic protein, myristoylated alanine-rich C kinase substrate (MARCKS), which has been implicated in regulating long-term neuroplastic events. Lenox and associates⁹⁵ demonstrated that chronic lithium administration dramatically reduced MARCKS expression in hippocampus, an effect that was not immediately reversed following lithium discontinuation. Subsequent studies carried out in immor-

Table 3. Signal Transduction Abnormalities in Bipolar Disorder: Preliminary Postmortem Findings ^a	
Increased $G\alpha_s$ immunoreactivity (but unchanged mRNA levels) Increased GTP γ S and forskolin-stimulated adenylyl cyclase activity Increased receptor/G protein coupling	
Increased $Ga_{q/1}$ levels and PKC β levels Reduced receptor-mediated phosphoinositide turnover	
Increased levels of selected PKC isozymes Reduced [³ H]cAMP binding (possible reduction in binding to regulatory subunits of PKA)	
Increased PKA activity	
^a Abbreviations: cAMP = cyclic AMP, G protein = guanine nucleotide binding protein, GTP = guanosine triphosphate, mRNA = messenger ribonucleic acid, PKA = protein kinase A, PKC = protein kinase C.	

talized hippocampal cells have demonstrated that this action of chronic lithium on MARCKS regulation is dependent on both the inositol concentration and the level of receptor-mediated activation of phosphoinositide hydrolysis.96 Recent studies provide evidence for the regulation of transcription as a major site for the action of chronic lithium on MARCKS expression in brain.⁹⁷ Since MARCKS may offer a specific target for pharmacologic agents to aim for in mood-stabilizing drugs, the effects of the anticonvulsant valproate as well as lithium were studied in immortalized hippocampal cells and were shown to induce a time- and concentration-dependent reduction in MARCKS protein expression. The activity of valproate was observed within a concentration range and time course considered consistent with clinical studies of patients with bipolar disorder. Additionally, therapeutic concentrations of combined lithium and valproate induced an additive reduction in MARCKS, consistent with experimental findings that the 2 drugs work through different mechanisms on the PKC system and with the clinical observation of the additivity of the 2 drugs in treatment responses.⁴

G Proteins

Several independent laboratories have examined G proteins in patients with mood disorders (Table 3).46,47,81,98 Young and associates^{46,99} were the first to report increased levels of $G\alpha_s$ in bipolar disorder patients in 2 separate studies. Compared with controls matched with respect to age, postmortem interval, and brain pH, they found increased levels of $G\alpha_{s}$ in frontal, temporal, and occipital cortices but not in hippocampus, thalamus, or cerebellum in postmortem brain tissue from patients with bipolar disorder.46,99 As discussed, this group also found increases in FSKstimulated adenylate cyclase activity in postmortem brain, compatible with a postreceptor abnormality in bipolar disorder. The findings of elevated $G\alpha_s$ levels and/or function are also supported by the recent findings of Friedman and Wang,100 who found increased agonist-activated $[^{35}S]GTP\gamma S$ binding to G protein α subunits in frontal cortical membrane preparations from bipolar disorder postmortem brain. Garcia-Sevilla and colleagues¹⁰¹ have reported increased levels of $G\alpha_{i1/2}$ in prefrontal cortical samples obtained at postmortem from depressed patients who committed suicide, effects that were apparently attenuated by antemortem antidepressant treatment. Overall, the findings in postmortem brain tissue in unipolar depression have been less consistent (see Warsh et al.47 for a summary of postmortem findings). In keeping with the G protein abnormalities in brain tissue obtained at postmortem from bipolar disorder patients, Schreiber and colleagues¹⁰² reported "hyperfunctional" G protein function in leukocytes of untreated manic patients by demonstrating that agonist-stimulated binding of [³H]Gpp(NH)pm (a stable, nonhydrolyzable analog of GTP) was enhanced in leukocyte membranes of untreated manic patients compared with controls. These findings suggest the presence of increased levels of G proteins and/or enhanced receptormediated activation of G proteins in leukocytes from untreated manic subjects. More recently, investigators have reported significantly higher levels of $G\alpha_s$ in mononuclear leukocytes from depressed bipolar but not unipolar patients.¹⁰³ Another study^{52,53} quantitated the levels of the major G protein α subunits in both platelet and leukocyte membranes from untreated (predominantly manic) and lithium-treated, euthymic bipolar disorder patients and observed higher levels of the 45 kDa form of $G\alpha$ in the combined group of bipolar disorder patients (treated and untreated) compared with controls. A recent study¹⁰⁴ has found elevated levels of Ga, mRNA in granulocytes obtained from bipolar but not unipolar patients. This study also found nonsignificant elevations in the levels of $G\alpha_{i2}$ in unmedicated bipolar patients, which, intriguingly, were modulated by lithium in bipolar but not unipolar patients. Similar to what has been observed in the CNS, one recent study¹⁰⁵ has evaluated the role of platelet G proteins as "signal coincidence detectors" and has found this function to be impaired in depressed patients. Overall, the most consistent finding to emerge is that in both peripheral cells and postmortem brain tissue from bipolar patients, elevations are observed in the predominant subspecies of $G\alpha_s$ present in the tissues examined. Since $G\alpha_s$ is a ubiquitously expressed protein, it may appear counterintuitive that an abnormality in this protein may play a role in the pathophysiology of bipolar disorder. However, there is already precedence for clinical disorders arising from abnormalities in the levels of $G\alpha_s$, which present with limited clinical manifestations, despite the ubiquitous expression of the protein.³⁹ These heterogeneous clinical effects and tissue-specific manifestations have been postulated to arise from differences in receptor, G protein, and effector stoichiometries in different tissues and from tissue-specific differences in the ability of different cells to compensate for the abnormality. It should be emphasized, however, that there is at present no evidence to suggest that the alterations in the levels of $G\alpha_s$ are due to a mutation in the $G\alpha_s$ gene itself.¹⁰⁶ Indeed, there are numerous transcriptional and posttranscriptional mechanisms that regulate the levels of G protein α subunits, and the elevated levels of $G\alpha_s$ could potentially represent the sequelae of alterations in any one of these other biochemical pathways. Thus, at this point, considerable caution is clearly required in the interpretation of the data, since they derive primarily from peripheral cell models and may not adequately reflect CNS pathology. The possibility of abnormalities in the biochemical pathways that regulate $G\alpha_s$ levels in bipolar disorder is currently undergoing further study.⁴⁷

Lithium and G proteins. Considerable evidence now indicates that chronic lithium administration affects G protein function^{3,40,41,63}; however, the preponderance of the data suggests that lithium, at therapeutically relevant concentrations, has no direct effects on G proteins and does not consistently alter G protein subunit levels.^{1,107} Although some studies have reported modest changes in the levels of G protein subunits, the preponderance of the data suggests that the effects of chronic lithium on signaltransducing properties occur in the absence of changes in the levels of G protein subunits per se.¹ Chronic in vivo lithium treatment has been shown to produce a significant increase in pertussis toxin-catalyzed [32P] adenosine diphosphate (ADP)-ribosylation in rat frontal cortex and human platelets. Since pertussis toxin selectively ADPribosylates the undissociated, inactive $\alpha\beta\gamma$ heterotrimeric form of G_i, these results suggest that lithium attenuates G_i function via a stabilization of the inactive conformation. These results suggest that the removal of the "inhibitory tone" by lithium may be responsible for the elevations in basal AC and the responses to agents activating the stimulatory pathway distal to the receptor.^{43,58} Consistent with this hypothesis, lithium has been shown to potentiate the hyperactivity induced by intra-accumbens cholera toxin administration (which activates the stimulatory G proteins, G_s and G_{olf}, ¹⁰⁸ These long-term effects of chronic lithium on G protein are most likely attributable to an indirect posttranslational modification of the G protein(s) and a relative change in the dynamic equilibrium of the active/ inactive states of protein conformation. In this context, it is noteworthy that investigators have demonstrated that lithium alters the levels of endogenous ADP-ribosylation in C6 glioma cells¹⁰⁹ and in rat brain,¹¹⁰ suggesting another mechanism by which chronic lithium may indirectly regulate the activity of these critical signaling proteins.

Valproate and G proteins. Recent studies have examined the effects of valproate on components of the β adrenergic receptor–coupled cAMP generating system.¹¹¹ Chronic valproate has been shown to produce a significant alteration of the β -adrenergic receptor–coupled cAMP generating system in cultured cells in vitro; these effects were observed at concentrations of valproate similar to those attained in the plasma in the clinical treatment of neuropsychiatric disorders. In contrast to what has been observed with chronic lithium treatment (discussed above), it was found that chronic valproate produced a significant reduction in the density of β -adrenergic receptors. Interestingly, the decrease in numbers of β -adrenergic receptors (approximately 30%) was accompanied by an even greater decrease in receptor- and postreceptor-mediated cAMP accumulation, suggesting that chronic valproate also exerts effects at the β -adrenergic receptor/G_s interaction or at postreceptor sites (e.g., G_s, AC). Consistent with such a contention, it was indeed found that chronic, but not acute, valproate incubation induced a marked decrease in the levels of $G\alpha_s$ 45 but no other G protein α subunits examined (G α_s 52, G $\alpha_{i1/2}$, G α_o , or G $\alpha_{a/11}$). In view of the suggested involvement of G_s in the pathophysiology of manic depressive illness (vide supra), as well as the effects of lithium on the β -adrenergic receptor/G/AC system, these effects may play a role in valproate's therapeutic effects and are worthy of further study.

Abnormalities of Calcium Signaling in Bipolar Disorder

Acting via intracellular proteins such as MARCKS and calmodulin, and enzymes such as PKC, AC, and CaM kinase, calcium ions have been shown to regulate the synthesis and release of neurotransmitters, neuronal excitability, cytoskeletal remodeling, and long-term neuroplastic events. Thus, it is not surprising that a large number of studies have investigated intracellular Ca²⁺ in peripheral cells in bipolar disorder.^{46,112,113} In view of the caveats associated with studies of peripheral circulating cells, the remarkable consistency of the findings is surprising indeed. Studies have consistently revealed elevations in both resting and stimulated intracellular Ca²⁺ levels in platelets, lymphocytes, and neutrophils of patients with bipolar disorder. The calcium abnormalities have been postulated to represent state-dependent findings,¹¹² but recent studies using transformed lymphoblasts from bipolar disorder patients have revealed similar abnormalities, suggesting that they may be trait dependent.¹¹³ The regulation of free intracellular Ca2+ is a complex, multifaceted process involving extracellular entry, release from intracellular stores following receptor-stimulated phosphoinositide hydrolysis, uptake into specific organelles, and binding to specific proteins. Thus, the abnormalities observed in bipolar disorder could arise from abnormalities at a variety of levels, and recent studies suggest that the abnormality lies beyond the receptor.¹¹⁴ Interestingly, recent studies have demonstrated that the Rap1 phosphorylation state is related to platelet intracellular calcium signaling,¹¹⁵ suggesting a possible relationship between these 2 documented abnormalities. Since PKC is also known to regulate calcium signaling at multiple levels, more recent studies have investigated the putative role of PKC in mediating calcium abnormalities in bipolar disorder. Preliminary analysis suggests that alterations in tonic PKC activity may play an important role in mediating the abnormal intracellular calcium responses observed in bipolar disorder (H.K.M., R. M. Post, M.D., unpublished observations, October 1996).

Do Animal Models Support the Involvement of Signaling Pathways in the Pathophysiology of Bipolar Disorder?

The use of animal models of disease and in vitro experiments to clarify complex neuropsychiatric disorders in humans has been reviewed, and caution must be exercised when applying results from these restricted models to the human environment.116,117 Two current models that have had reasonable heuristic value in the study of mood disorders have been kindling and behavioral sensitization.^{116,117} The increased, sensitized behavioral response, measured through behavioral or electrophysiologic changes to repetition of the same stimulus, whether a pharmacologic agent or an electrical challenge, can persist for months or years after discontinuation of the stimuli. The cellular mechanisms responsible for these long-lasting behavioral responses in kindled or stimulantsensitized animals are actively under study, including both G protein-mediated pathways and PKC signaling. Briefly, evidence that regulation of G proteins is a component of the biochemical alterations inherent in the effects of chronic stimulation comes from the significant enhancement of psychostimulant-induced motor activity and release of dopamine in the nucleus accumbens reported after treatment with pertussis toxin, an inhibitor of specific G proteins (G_i and G_o).¹¹⁸ Additionally, specific decreases in G protein subunit concentrations in the ventral tegmental area and nucleus accumbens of rats have been reported in response to chronic cocaine administration.¹¹⁹

Alterations in PKC function have been implicated through experiments with chronic and acute stimulant administration leading to sensitization and kindling as well as alterations in neurotransmitter release during amphetamine administration in animal models. Substantial increases in membrane-associated but not cytosolic PKC have been documented in the bilateral hippocampus for up to 4 weeks and in the amygdala/pyriform cortex at 4 weeks as well as after the last seizure in hippocampal-kindled rats.¹²⁰ Studies of perfused rat striatial slices treated with PKC stimulators or inhibitors also show that amphetamine-mediated dopamine release through the plasmalemmal dopamine transporter is dependent on PKC function.¹²¹ Since drug treatment with either lithium or valproate also targets components of the PKC pathway with a diametrically opposite effect in critical limbic structures in behaviorally sensitized and kindled rats, one has greater confidence in extrapolating these results to humans.42 Extensive data also suggest that long-term alterations in midbrain dopaminergic neurotransmission are involved in the establishment of behavioral sensitization, and recent data implicate a cascade of changes in gene expression, including neurotropic and apoptotic factors.¹¹⁶ Therefore, induced behaviors established in animal models now provide experimental systems in which to explore the contributions of various signal transduction pathways and gene control elements.

CIRCADIAN RHYTHMS

Mood disorders are intimately associated with disruptions in circadian rhythms; this may be particularly true for bipolar disorder, which is characterized by an episodic pattern, hallmark recurrence, and fluctuations in symptoms and severity, course of illness, and responses to treatments. Depressed patients frequently evidence shortened rapid eye movement latency, early wakening, and advances in hormonal and temperature rhythms, implying dysregulation of internal oscillators with respect to other internal clock systems or external time-dependent stimuli.5,122,123 A circadian clock located in the hypothalamic suprachiasmatic nucleus regulates the circadian phases of mammalian locomotor activity, which during synchronization can cause time-consistent circadian rhythms of very distinct amplitudes. Studies on model systems in Drosophila mutants have located circadian clock genes that are strongly suspected, through additional studies with mouse genes, to be highly evolutionarily conserved.¹²⁴ Since chronic (not acute) lithium treatment affects, although with variabilities in advancing or delaying the time of highest amplitude, all biological systems in an evolutionarily conserved manner, genes controlling the circadian clock might be awry in mood disorders and be the ultimate targets of lithium.¹²⁵ Without question, these studies are in their infancy and must be further supported with additional investigations.111,126

CELL DEATH AND ATROPHY IN MOOD DISORDERS

Studies have recently demonstrated that although mood disorders have traditionally been conceptualized as neurochemical disorders, reductions in regional CNS volume and cell numbers (both neurons and glia) are observed in many patients. One line of evidence comes from structural imaging studies, which have recently begun to provide important clues about the neuroanatomical basis of mood disorders. Although the findings are not as consistent as those reported in schizophrenia, in toto, the volumetric neuroimaging studies have demonstrated an enlargement of third and lateral ventricles, as well as reduced gray matter volumes in parts of the orbital and medial prefrontal cortices, the ventral striatum, and the mesiotemporal cortex.^{21,127} Lending support to the structural neuroimaging literature are multiple functional brain imaging studies that have shown abnormalities in metabolic rate and blood flow in these same areas in major depression.²¹ In addition to the accumulating neuroimaging evidence, several postmortem brain studies are now providing direct evidence for reductions in regional CNS volume and cell number.^{42,43,128} Together, the preponderance of the data from the neuroimaging studies and the growing body of postmortem evidence presents a convincing case that there is indeed a reduction in regional CNS volume, accompanied by a reduction in cell numbers in at least a subset of patients with mood disorders. It is thus noteworthy that recent studies have led to a completely unexpected target for the actions of chronic lithium and valproate in the frontal cortex—the cytoprotective protein bcl-2.^{2,129}

Chronic treatment of rodents with "therapeutic" doses of lithium and valproate was found to produce a *doubling* of bcl-2 levels in frontal cortex, an effect that was primarily due to a marked increase in the number of bcl-2 immunoreactive cells in layers II and III of frontal cortex. Interestingly, the importance of neurons in layers II-IV of the frontal cortex in mood disorders has recently been emphasized, since primate studies have indicated that these are important sites for connections with other cortical regions and major targets for subcortical input.¹³⁰ Furthermore, these are the very same brain areas where the greatest neuronal or glial diminution has been observed in postmortem studies of major depression subjects.^{128,129} Chronic lithium also markedly increased the number of bcl-2 immunoreactive cells in the dentate gyrus and striatum.² The lithiuminduced increase in bcl-2 levels has also been convincingly replicated in rat cerebellar granule cells in recent studies.¹³⁰ Consistent with the known cytoprotective effects of bcl-2, lithium, at therapeutically relevant concentrations, has been shown to exert neuroprotective effects in a variety of paradigms. Thus, in vitro, lithium has been demonstrated to protect against the deleterious effects of glutamate, N-methyl-p-aspartate receptor activation, aging, serum/nerve growth factor deprivation, ouabain, and thapsigargin (which mobilizes intracellular 1-methyl-4phenypyradinium ion [MPP⁺], Ca²⁺, and β -amyloid).^{2,43,44}

Cytoprotective effects of lithium have also been demonstrated in rodent brain in vivo. Thus, lithium treatment has been shown to attenuate the biochemical deficits produced by kainic acid infusion, ibotenic acid infusion, and forebrain cholinergic system lesions and to also attenuate the behavioral deficits produced by forebrain cholinergic system lesions.131,132 Recent studies by Chuang and associates have shown that lithium exerts dramatic protective effects against middle cerebral artery occlusion, reducing not only the infarct size (56%) but also the neurologic deficits (abnormal posture and hemiplegia).¹³³ Most recently, the same research group has demonstrated that in vivo lithium treatment robustly protects neurons in the striatum from quinolinic acid-induced toxicity in a putative model of Huntington's disease.¹³⁴ It is indeed striking that this seemingly simple monovalent cation that has been used clinically for decades has been shown to exert neuroprotective effects in these preclinical models.

In addition to its effects on bcl-2, the effects of lithium on other signaling pathways and transcription factors40,41,52,53 may also contribute to its neuroprotective effects. Perhaps foremost among these is glycogen synthase kinase-3 β (GSK-3 β). GSK-3 β is an evolutionarily highly conserved enzyme known to play a critical role in development as well as in the mature CNS by regulating various cytoskeletal events and gene expression. It is now clear that lithium, at concentrations similar to those attained clinically, is an inhibitor of GSK-36.135 Since GSK-36 has been demonstrated to contribute to apoptotic cell death¹³⁶ and appears to play a role in the hyperphosphorylation of tau observed in Alzheimer's disease, there has been considerable recent excitement about the possibility of developing novel GSK-3β inhibitors not only for the treatment of recurrent major depression, but potentially also for more classic neurodegenerative diseases.^{40,41,43}

Does Lithium Affect Neurogenesis?

The demonstration that neurogenesis occurs into senescence in the human brain has generated considerable excitement about the possibility of pharmacologically treating disease-related neuronal atrophy or loss.137,138 The localization of pluripotent progenitor cells and neurogenesis occurs in restricted brain regions. The greatest density of new cell birth is observed in the subventricular zone and the subgranular layer of the hippocampus. Cells born in the subventricular zone migrate largely to the olfactory bulb, and those born in the subgranular zone migrate into the granule cell layer. A large number of the newborn daughter cells are known to die rapidly, likely via apoptosis.138 Thus, increasing bcl-2 levels could enhance the survival of the newborn cells, allowing them to differentiate into neurons. Additionally, bcl-2 has been shown to have robust effects on the regeneration of CNS axons.¹³⁹ In view of bcl-2's major neuroprotective and neurotrophic role, a study⁹¹ was undertaken to determine if lithium, administered at therapeutically relevant concentrations, affects neurogenesis in the adult rodent brain. (Kempermann and Gage¹³⁸ have suggested the use of neurogenesis to refer to a series of events [including proliferation of a neuronal precursor or stem cell and survival of the daughter cells] that result in the appearance of a new neuron.) To investigate the effects of chronic lithium administration on neurogenesis, mice were treated with "therapeutic" lithium (mean \pm SD plasma levels = 0.97 \pm 0.20 mM) for ~4 weeks. After treatment with lithium for 14 days, the mice were administered single doses of BrdU (bromodeoxyuridine, a thymidine analog that is incorporated into the DNA of dividing cells) for 12 consecutive days. Lithium treatment continued throughout the duration of the BrdU administration. Following BrdU immunohistochemistry, 3-dimensional cell counting was performed using a computer-assisted image analysis system. This system is based on the optical disector method and estimates the

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number of cells, independent of section thickness and cell shape. It was found that chronic lithium administration does, indeed, result in an increase in the number of BrdU-positive cells in the dentate gyrus.⁹¹ Moreover, approximately two thirds of the BrdU-positive cells also double-stained with the neuronal marker NeuN, confirming their neuronal identity. Double-staining of BrdU and bcl-2 was also observed, and studies using bcl-2 transgenic animals are currently under way to delineate the role of bcl-2 over-expression in mediating the effects of lithium on hip-pocampal neurogenesis.

Can Lithium's Neurotrophic and Neuroprotective Effects be Demonstrated Longitudinally in the Human Brain in Vivo?

Although a considerable body of preclinical data now demonstrates neurotrophic and neuroprotective effects of lithium in preclinical models, it has not been clear if lithium can actually exert similar neurotrophic effects in the human brain in vivo. A longitudinal clinical study¹⁴⁰ was undertaken, using proton MRS to quantitate N-acetylaspartate (NAA) levels. NAA has been proposed to represent a putative marker of neuronal viability and has been utilized to follow the course of neurodegenerative disorders. It was found that chronic lithium administration at therapeutic doses increased NAA concentration in the human brain in vivo. Furthermore, a striking correlation (~0.97) was observed between lithium-induced NAA increases and regional voxel gray matter content. A followup study¹⁴¹ was undertaken to determine if lithium-induced increases in bcl-2 would also lead to neuronal increases, and thus to increased brain gray matter volume in patients with manic depressive illness. Brain tissue volumes were determined using high-resolution 3-dimensional MRI at baseline (medication-free, after a minimum 14-day washout), and then repeated after 4 weeks of lithium at therapeutic doses. This study showed that chronic lithium significantly increases total gray matter content in the human brain of patients with manic depressive illness. No significant changes were observed in brain white matter volume or in quantitative measures of regional cerebral water content, thereby providing strong evidence that the observed increases in gray matter content are likely owing to neurotrophic effects as opposed to any possible cell swelling or osmotic effects associated with lithium treatment.

CONCLUSION

As we have discussed, the complexity of the biological and behavioral features of mania and depression has resulted in a vast, diverse compendium of corresponding data that have contributed to the understanding of the pathophysiology of bipolar disorder at different biochemical and neurobiological levels. Although preclinical models and clinical studies are challenging to design so that they mirror the variability of the illness in terms of its acute and chronic nature, episodic and cycling clinical status, and associated morbidity, fairly robust results have been obtained for a variety of systems. Neurotransmitter dysfunction has been confirmed in biochemical, physiologic, and neuroanatomical studies in patients and animal models and has become a target for pharmacologic intervention. Abnormalities in signal transduction cascades are also strongly supported in the underlying neurobiology of the disorder because of their pivotal biological role in areas of the brain associated with the disease process, their similarity with known diseases with etiologic aberrant signaling function, and their experimental findings during drug modulation. More recently, application of new investigational approaches to study the temporal and spatial pattern of gene expression changes has revealed a potential role for neurotrophic and neuroprotective factors both in the pathophysiology of disease progression and in optimal long-term treatment by mood stabilizers.40,44,142

Drug names: carbamazepine (Tegretol and others), clozapine (Clozaril and others), lamotrigine (Lamictal), olanzapine (Zyprexa), risperidone (Risperdal), tamoxifen (Nolvadex), topiramate (Topamax).

REFERENCES

- Lenox RH, Manji HK. Lithium. In: Nemeroff CB, Schatzberg AF, eds. American Psychiatric Press Textbook of Psychopharmacology. 2nd ed. Washington, DC: American Psychiatric Press; 1998:379–430
- Manji HK, Moore GJ, Chen G. Lithium at 50: have the neuroprotective effects of this unique cation been overlooked? Biol Psychiatry 1999;46: 929–940
- Manji HK, Lenox RH. Signaling: cellular insights into the pathophysiology of bipolar disorder. Biol Psychiatry 2000;48:518–530
- Watson DG, Watterson JM, Lenox RH. Sodium valproate down-regulates the myristoylated alanine-rich C kinase substrate (MARCKS) in immortalized hippocampal cells: a property of protein kinase C-mediated mood stabilizers. J Pharmacol Exp Ther 1998;285:307–316
- Goodwin FK, Jamison KR. Manic-Depressive Illness. New York, NY: Oxford University Press; 1990
- Goodwin F, Ghaemi N. Bipolar disorder: state of the art. Dialogues Clin Neurosci 1999;1:41–51
- Schildkraut JJ. The catecholamine hypothesis of affective disorders: a review of the supporting evidence. Am J Psychiatry 1965;22:509–522
- Bunney WE, Davis JM. Norepinephrine in depressive reactions: a review. Arch Gen Psychiatry 1965;13:483–494
- Janowsky DS, Overstreet DH. The role of acetylcholine mechanisms in mood disorders. In: Bloom FE, Kupfer DJ, eds. Psychopharmacology: The Fourth Generation of Progress. New York, NY: Raven Press; 1995: 945–956
- Janowsky DS, El-Yousef MK, Davis JM, et al. A cholinergic-adrenergic hypothesis of mania and depression. Lancet 1972;2:632–635
- Racagni G, Brunello N. Physiology to functionality: the brain and neurotransmitter activity. Int Clin Psychopharmacol 1999;14(suppl 1):S3–S7
- Wehr TA, Goodwin FK. Can antidepressants cause mania and worsen the course of affective illness? Am J Psychiatry 1987;144:1403–1411
- Manji HK, Potter WZ. Monoaminergic mechanisms in bipolar disorder. In: Young LT, Joffe RT, eds. Biological Models and Their Clinical Application. New York, NY: Marcel Dekker; 1997:1–40
- Masand PS, Gupta S. Selective serotonin-reuptake inhibitors: an update. Harv Rev Psychiatry 1999;7:69–84
- Schatzberg AF, Schildkraut JJ. Recent studies on norepinephrine systems in mood disorders. In: Bloom FE, Kupfer DJ, eds. Psychopharmacology: The Fourth Generation of Progress. New York, NY: Raven Press; 1995: 911–920
- 16. Potter WZ, Rudorfer MV, Goodwin FK. Bipolar Disorder. In: Hales RE,

Frances AJ, eds. American Psychiatric Association Annual Review, vol 6. Washington, DC: American Psychiatric Press; 1987:32–60

- Kopin IJ. Catecholamine metabolism: basic aspects and clinical significance. Pharmacol Rev 1985;37:333–364
- Bowden CL. Toward an integrated biological model of bipolar disorder. In: Young LT, Joffe RT, eds. Bipolar Disorder: Biological Models and Their Clinical Application. New York, NY: Marcel Dekker; 1997:235–254
- Maes M, Meltzer HY, D'Hondt P, et al. Effects of serotonin precursors on the negative feedback effects of glucocorticoids on hypothalamic-pituitaryadrenal axis function in depression. Psychoneuroendocrinology 1995;20: 149–167
- Garlow SJ, Musselman DL, Nemeroff CB. The neurochemistry of mood disorders: clinical studies. In: Charney DS, Nester EJ, Bunney BS, eds. Neurobiology of Mental Illness. New York, NY: Oxford University Press; 1999:348–364
- Drevets WC, Frank E, Price JC, et al. PET imaging of serotonin 1A receptor binding in depression. Biol Psychiatry 1999;46:1375–1387
- Delgado PL, Miller HL, Salomon RM, et al. Tryptophan-depletion challenge in depressed patients treated with desipramine or fluoxetine: implications for the role of serotonin in the mechanism of antidepressant action. Biol Psychiatry 1999;46:212–220
- Cappiello A, Sernyak MJ, Malison RT, et al. Effects of acute tryptophan depletion in lithium-remitted manic patients: a pilot study. Biol Psychiatry 1997;42:1076–1078
- Cassidy F, Murry E, Carroll BJ. Tryptophan depletion in recently manic patients treated with lithium. Biol Psychiatry 1998;43:230–232
- 25. Fibiger HC. The dopamine hypothesis of schizophrenia and depression: contradictions and speculations. In: Willner P, Scheel-Kruger H, eds. The Mesolimbic Dopamine System: From Motivation to Action. Chichester, England: Wiley Press; 1991:615–637
- Willner P. Dopaminergic mechanisms in depression and mania. In: Bloom FE, Kupfer DJ, eds. Psychopharmacology: The Fourth Generation of Progress. New York, NY: Raven Press; 1995:921–931
- Goodwin FK, Sack RL. Central dopamine function in affective illness: evidence from precursors, enzyme inhibitors, and studies of central dopamine turnover. Adv Biochem Psychopharmacol 1974;12:261–279
- 28. Swann AC, Katz MM, Bowden CL. Psychomotor performance and monoamine function in bipolar and unipolar affective disorders. Biol Psychiatry 1999:45:979–988
- 29. Berman RM, Narasimhan M, Miller HL, et al. Transient depressive relapse induced by catecholamine depletion: potential phenotypic vulnerability marker? Arch Gen Psychiatry 1999;56:395–403
- Anand A, Darnell A, Miller HL, et al. Effect of catecholamine depletion on lithium-induced long-term remission of bipolar disorder. Biol Psychiatry 1999;45:972–978
- Holsboer F. Neuroendocrinology of mood disorders. In: Bloom FE, Kupfer DJ, eds. Psychopharmacology: The Fourth Generation of Progress. New York, NY: Raven Press; 1995:957–969
- Barden N, Reul JM, Holsboer F. Do antidepressants stabilize mood through actions on the hypothalamic-pituitary-adrenocortical system? Trends Neurosci 1995;18:6–11
- Sapolsky RM. Stress, glucocorticoids, and damage to the nervous system: the current state of confusion. Stress 1996;1:1–19
- McEwen BS. Stress and hippocampal plasticity. Ann Rev Neurosci 1999;22:105–122
- Bhalla US, Iyengar R. Emergent properties of networks of biological signaling pathways. Science 1999;283:381–387
- Bourne HR, Nicoll R. Molecular machines integrate coincident synaptic signals. Cell Suppl 1993;72:65–75
- Weng G, Bhalla US, Iyengar R. Complexity in biological signaling systems. Science 1999;284:92–96
- Milligan G, Wakelam M. G Proteins: Signal Transduction and Disease. San Diego, Calif: Academic Press; 1992
- Spiegel AM. G Proteins, Receptors, and Disease. Totowa, NJ: Humana Press; 1998
- Jope RS. Anti-bipolar therapy: mechanism of action of lithium. Mol Psychiatry 1999;4:117–128
- Jope RS. A bimodal model of the mechanism of action of lithium. Mol Psychiatry 1999;4:21–25
- Manji HK, Lenox RH. Protein kinase C signaling in the brain: molecular transduction of mood stabilization in the treatment of bipolar disorder. Biol Psychiatry 1999;46:1328–1351
- 43. Manji HK, Chen G, Hsiao JK, et al. Regulation of signal transduction path-

ways by mood stabilizing agents: implications for the pathophysiology and treatment of bipolar affective disorder. In: Manji HK, Bowden CL, Belmaker RH, eds. Bipolar Medications: Mechanisms of Action. Washington, DC: American Psychiatric Press; 2000:129–177

- 44. Manji HK, Moore GJ, Chen G. Clinical and preclinical evidence for the neurotrophic effects of mood-stabilizing agents: implications for the pathophysiology and treatment of manic-depressive illness. Biol Psychiatry. In press
- Lenox RH. Role of receptor coupling to phosphoinositide metabolism in the therapeutic action of lithium. Adv Exp Med Biol 1987;221:515–530
- 46. Wang JF, Young LT, Li PP, et al. Signal transduction abnormalities in bipolar disorder. In: Young LT, Joffe RT, eds. Bipolar Disorder: Biological Models and Their Clinical Application. New York, NY: Marcel Dekker; 1997:41–79
- Warsh JJ, Young LT, Li PP. Guanine nucleotide binding (G) protein disturbances. In: Manji HK, Bowden CL, Belmaker RH, eds. Bipolar Affective Disorder in Bipolar Medications: Mechanisms of Action. Washington, DC: American Psychiatric Press; 2000:299–329
- Perez J, Tardito D, Mori S, et al. Abnormalities of cAMP signaling in affective disorders: implications for pathophysiology and treatment. Bipolar Disord 2000;2:27–36
- Perez J, Tardito D, Mori S, et al. Altered Rap1 endogenous phosphorylation and levels in platelets from patients with bipolar disorder. J Psychiatr Res 2000;34:99–104
- Rahman S, Li PP, Young LT, et al. Reduced [3H]cyclic AMP binding in postmortem brain from subjects with bipolar affective disorder. J Neurochem 1997;68:297–304
- Fields A, Li PP, Kish SJ, et al. Increased cyclic AMP-dependent protein kinase activity in postmortem brain from patients with bipolar affective disorder. J Neurochem 1999;73:1704–1710
- Manji HK, Chen G, Shimon H, et al. Guanine nucleotide binding proteins in bipolar affective disorder: effects of long-term lithium treatment. Arch Gen Psychiatry 1995;52:135–144
- Manji HK, Potter WZ, Lenox RH. Signal transduction pathways: molecular targets for lithium's actions. Arch Gen Psychiatry 1995;52:531–543
- Mork A, Geisler A, Hollund P. Effects of lithium on second messenger systems in the brain. Pharmacol Toxicol 1992;71(suppl 1):4–17
- Andersen PH, Geisler A. Lithium inhibition of forskolin-stimulated adenylate cyclase. Neuropsychobiology 1984;12:1–3
- Mork A, Geisler A. The effects of lithium in vitro and ex vivo on adenylate cyclase in brain are exerted by distinct mechanisms. Neuropharmacology 1989;28:307–311
- Newman ME, Belmaker RH. Effects of lithium in vitro and ex vivo on components of the adenylate cyclase system in membranes from the cerebral cortex of the rat. Neuropharmacology 1987;26:211–217
- Masana MI, Bitran JA, Hsiao JK, et al. In vivo evidence that lithium inactivates Gi modulation of adenylate cyclase in brain. J Neurochem 1992; 59:200–205
- Wiborg O, Kruger T, Jakobsen SN. Region-selective effects of long-term lithium and carbamazepine administration on cyclic AMP levels in rat brain. Pharmacol Toxicol 1999;84:88–93
- Colin SF, Chang HC, Mollner S, et al. Chronic lithium regulates the expression of adenylate cyclase and Gi-protein alpha subunit in rat cerebral cortex. Proc Natl Acad Sci U S A 1991;88:10634–10637
- Jensen JB, Mikkelsen JD, Mork A. Increased adenylyl cyclase type 1 mRNA, but not adenylyl cyclase type 2 in the rat hippocampus following antidepressant treatment. Eur Neuropsychopharmacol 2000;10:105–111
- Ozaki N, Manji HK, Luiberman V, et al. A naturally occurring amino acid substitution of the human serotonin (5-HT) 2A receptor influences amplitude and timing of intracellular calcium mobilization. J Neurochemistry 1997;68:2186–2193
- Wang HY, Friedman E. Effects of lithium on receptor-mediated activation of G proteins in rat brain cortical membranes. Neuropharmacology 1999;38:403–414
- Wang JF, Asghari V, Rockel C, et al. Cyclic AMP responsive element binding protein phosphorylation and DNA binding is decreased by chronic lithium but not valproate treatment of SH-SY5Y neuroblastoma cells. Neuroscience 1999;91:771–776
- 65. Risby ED, Hsiao JK, Manji HK, et al. The mechanisms of action of lithium. Arch Gen Psychiatry 1991;48:513–524
- Post RM, Weiss SR, Chuang DM. Mechanisms of action of anticonvulsants in affective disorders: comparisons with lithium. J Clin Psychopharmacol 1992;12:23S–35S

- Post RM, Weiss SRB, Clark M, et al. Lithium, carbamazepine, and valproate in affective illness: biochemical and neurological mechanisms. In: Manji HK, Bowden CL, Belmaker RH, eds. Bipolar Medications: Mechanisms of Action. Washington, DC: American Psychiatric Press; 2000: 219–248
- Palmer GC, Jones DJ, Medina MA, et al. Anticonvulsant drug actions on in vitro and in vivo levels of cyclic AMP in the mouse brain. Epilepsia 1979;20:95–104
- Palmer GC. Interactions of antiepileptic drugs on adenylate cyclase and phosphodiesterases in rat and mouse cerebrum. Exp Neurol 1979;63: 322–335
- Myllyla VV. Effect of convulsions and anticonvulsive drugs on cerebrospinal fluid cyclic AMP in rabbit. Eur Neurol 1976;4:97–107
- Elphick M, Taghavi Z, Powell T, et al. Chronic carbamazepine down-regulates adenosine A2 receptors: studies with the putative selective adenosine antagonists PD115,199 and PD116,948. Psychopharmacology (Berl) 1990; 100:522–529
- van Calker D, Steber R, Klotz KN, et al. Carbamazepine distinguishes between adenosine receptors that mediate different second messenger responses. Eur J Pharmacol 1991;206:285–290
- Lewin E, Bleck V. Cyclic AMP accumulation in cerebral cortical slices: effect of carbamazepine, phenobarbital, and phenytoin. Epilepsia 1977;18: 237–242
- Ferrendelli JA, Kinscherf DA. Inhibitory effects of anticonvulsant drugs on cyclic nucleotide accumulation in brain. Ann Neurol 1979;5:533–538
- 75. Divish MM, Sheftel G, Boyle A, et al. Differential effect of lithium on fos protooncogene expression mediated by receptor and postreceptor activators of protein kinase C and cyclic adenosine monophosphate: model for its antimanic action. J Neurosci Res 1991;28:40–48
- Chen G, Hawver D, Wright C, et al. Attenuation of adenylate cyclases by carbamazepine in vitro. J Neurochemistry 1996;67:2079–2086
- Sheng M, Greenberg ME. The regulation and function of c-fos and other immediate early genes in the nervous system. Neuron 1990;4:477–485
- Brown AS, Mallinger AG, Renbaum LC. Elevated platelet membrane phosphatidylinositol-4,5-bisphosphate in bipolar mania. Am J Psychiatry 1993;150:1252–1254
- 79. Soares JC, Dippold CS, Mallinger AG. Platelet membrane phosphatidylinositol-4,5-bisphosphate alterations in bipolar disorder: evidence from a single case study. Psychiatry Res 1997;69:197–202
- 80. van Calker D, Forstner U, Bohus M, et al. Increased sensitivity to agonist stimulation of the Ca²⁺ response in neutrophils of manic depressive patients; effect of lithium therapy. Neuropsychobiology 1993;27:180–183
- Mathews SR, Li PP, Young LT, et al. Increased G alpha q/11 immunoreactivity in postnortem occipital cortex from patients with bipolar affective disorder. Biol Psychiatry 1997;41:649–656
- Jope RS, Song L, Li PP, et al. The phosphoinositide signal transduction system is impaired in bipolar affective disorder brain. J Neurochem 1996; 66:2402–2409
- Friedman E, Hoau-Yan-Wang, Levinson D, et al. Altered platelet protein kinase C activity in bipolar affective disorder, manic episode. Biol Psychiatry 1993;33:520–525
- Wang HY, Friedman E. Enhanced protein kinase C activity and translocation in bipolar affective disorder brains. Biol Psychiatry 1996;40:568–575
- Hahn CG, Friedman E. Abnormalities in protein kinase C signaling and the pathophysiology of bipolar disease. Bipolar Disorders 1999;1:81–86
- Pandey GN, Dwivedi Y, Pandey SC, et al. Protein kinase C in the postmortem brain of teenage suicide victims. Neurosci Lett 1997;228:111–114
- Jope RS, Williams MB. Lithium and brain signal transduction systems. Biochem Pharm 1994;77:429–441
- Moore GJ, Bebchuk JM, Parrish JK, et al. Temporal dissociation between lithium-induced CNS *myo*-inositol changes and clinical response in manicdepressive illness. Am J Psychiatry 1999;156:1902–1908
- Chen G, Masana M, Manji HK. Lithium regulates PKC-mediated intracellular cross-talk and gene expression in the CNS in vivo. Bipolar Disord. In press
- Manji HK, Etcheberrigaray R, Chen G, et al. Lithium decreases membraneassociated protein kinase C in hippocampus: selectivity for the α isozyme. J Neurochem 1993;61:2303–2310
- 91. Chen G, Rajkowska G, Du F, et al. Enhancement of hippocampal neurogenesis by lithium. J Neurochem. 2000;75:1729–1734
- Li X, Jope RS. Selective inhibition of the expression of signal transduction proteins by lithium in nerve growth factor-differentiated PC12 cells. J Neurochem 1995;65:2500–2508

- Chen G, Manji HK, Hawver DB, et al. Chronic sodium valproate selectively decreases protein kinase C alpha and epsilon in vitro. J Neurochem 1994;63:2361–2365
- Bebchuk JM, Arfken CL, Dolan-Manji S, et al. A preliminary investigation of a protein kinase C inhibitor (tamoxifen) in the treatment of acute mania. Arch Gen Psychiatry 2000;57:95–97
- Lenox RH, Watson DGJP, Ellis J. Chronic lithium administration alters a prominent PKC substrate in rat hippocampus. Brain Res 1992;570: 333–340
- Watson DG, Lenox RH. Chronic lithium-induced down-regulation of MARCKS in immortalized hippocampal cells: potentiation by muscarinic receptor activation. J Neurochem 1996;67:767–777
- Wang L, Watterson JM, Watson DG, et al. Myristoylation regulates retinoic acid-induced down-regulation of MARCKS in transfected hippocampal cells. To be presented at the annual meeting of the Society for Neuroscience; Nov 4–9, 2000; New Orleans, La
- Chen G, Hasanat KA, Bebchuk JM, et al. Regulation of signal transduction pathways and gene expression by mood stabilizers and antidepressants. Psychosom Med 1999;61:599–617
- Young LT, Li PP, Kish SJ, et al. Cerebral cortex G_s α protein levels and forskolin-stimulated cyclic AMP formation are increased in bipolar affective disorder. J Neurochem 1993;61:890–898
- Friedman E, Wang HY. Receptor-mediated activation of G proteins is increased in postmortem brains of bipolar affective disorder subjects. J Neurochem 1996;67:1145–1152
- 101. Garcia-Sevilla JA, Escriba PV, Ozaita A, et al. Up-regulation of immunolabeled alpha2A-adrenoceptors, Gi coupling proteins, and regulatory receptor kinases in the prefrontal cortex of depressed suicides. J Neurochem 1999;72:282–291
- Schreiber G, Avissar S, Danon A, et al. Hyperfunctional G proteins in mononuclear leukocytes of patients with mania. Biol Psychiatry 1991; 29:273–280
- 103. Young LT, Li PP, Kamble A, et al. Mononuclear leukocyte levels of G proteins in depressed patients with bipolar disorder or major depressive disorder. Am J Psychiatry 1994;151:594–596
- Spleiss O, van Calker D, Scharer L, et al. Abnormal G protein alpha(s)and alpha(i2)-subunit mRNA expression in bipolar affective disorder. Mol Psychiatry 1998;3:512–520
- 105. Mooney JJ, Samson JA, McHale NL, et al. Signal transduction by platelet adenylate cyclase: alterations in depressed patients may reflect impairment in the coordinated integration of cellular signals (coincidence detection). Biol Psychiatry 1998;43:574–583
- 106. Ram A, Guedj F, Cravchik A, et al. No abnormality in the gene for the G protein stimulatory α subunit in patients with bipolar disorder. Arch Gen Psychiatry 1997;54:44–49
- Ellis J, Lenox RH. Receptor coupling to G proteins: interactions not affected by lithium. Lithium 1991;2:141–147
- Kofman O, Li PP, Warsh JJ. Lithium, but not carbamazepine, potentiates hyperactivity induced by intra-accumbens cholera toxin. Pharmacol Biochem Behav 1998;59:191–200
- Young LT, Woods CM. Mood stabilizers have differential effects on endogenous ADP ribosylation in C6 glioma cells. Eur J Pharmacol 1996; 309:215–218
- Nestler EJ, Terwilliger RZ, Duman S. Regulation of endogenous ADPribosylation by acute and chronic lithium in rat brain. J Neurochem 1995;64:2319–2324
- Chen G, Manji HK, Wright CB, et al. Effects of valproic acid on βadrenergic receptors, G-proteins and adenylate-cyclase in rat C6 glioma cells. Neuropsychopharmacology 1996;15:271–280
- Dubovsky S, Murphy J, Christiano J, et al. The calcium second messenger system in bipolar disorders: data supporting new research directions. J Neuropsychiatry 1992;4:3–14
- 113. Emanghoreishi M, Schlichter L, Li PP, et al. High intracellular calcium concentrations in transformed lymphoblasts from subjects with bipolar I disorder. Am J Psychiatry 1997;154:976–982
- 114. Hough C, Lu SJ, Davis CL, et al. Elevated basal and thapsigargin stimulated intracellular calcium of platelets and lymphocytes from bipolar affective disorder patients measured by a fluorometric microassay. Biol Psychiatry 1999;46:247–255
- 115. Magnier C, Corvazier E, Aumont MC, et al. Relationship between Rap1 protein phosphorylation and regulation of Ca²⁺ transport in platelets: a new approach. Biochem J 1995;310(pt 2):469–475
- 116. Post RM, Weiss SRB. Neurobiological models of recurrence in mood dis-

order. In: Charney DS, Nester EJ, Bunney BS, eds. Neurobiology of Mental Illness. New York, NY: Oxford University Press; 1999:365–384

- 117. Einat H, Kofman O, Belmaker RH. Animal models of bipolar disorder: from a single episode to progressive cycling models. In: Myslobodsky MS, Weiner I, eds. Contemporary Issues in Modeling Psychopathology. In press
- Steketee JD, Kalivas PW. Sensitization to psychostimulants and stress after injection of pertussis toxin into the A10 dopamine region. J Pharmacol Exp Ther 1991;259:916–924
- Nestler EJ, Terwilliger RZ, Walker JR, et al. Chronic cocaine treatment decreases levels of G protein subunits Gi alpha and Go alpha in discrete regions of rat brain. J Neurochem 1990;55:1079–1082
- Daigen A, Akiyama K, Otsuki S. Long-lasting change in the membraneassociated protein kinase C activity in the hippocampal kindled rat. Brain Res 1991;545:131–136
- Kantor L, Gnegy ME. Protein kinase C inhibitors block amphetaminemediated dopamine release in rat striatal slices. J Pharmacol Exp Ther 1998;284:592–598
- Wirz-Justice A. Biological rhythms in mood disorders. In: Bloom FE, Kupfer DJ, eds. Psychopharmacology: The Fourth Generation of Progress. New York, NY: Raven Press; 1995:999–1017
- 123. Wehr TA, Goodwin FK. Biological rhythms in manic-depressive illness. In: Wehr TA, Goodwin FK, eds. Circadian Rhythms in Psychiatry. Pacific Grove, Calif: Boxwood Press; 1983:129–184
- King DP, Takahashi JS. Molecular genetics of circadian rhythms in mammals. Ann Rev Neurosci 2000;22:712–742
- 125. Bunney WE, Bunney BG. Molecular clock genes in man and lower animals: possible implications for circadian abnormalities in depression. Neuropsychopharmacology 2000;22:225–245
- Ikonomov O, Manji HK. Molecular mechanisms underlying moodstabilization in manic-depressive illness: the phenotype challenge. Am J Psychiatry 1999;156:1506–1514
- Soares JC, Mann JJ. The anatomy of mood disorders: review of structural neuroimaging studies. Biol Psychiatry 1997;41:86–106
- 128. Rajkowska G. Postmortem studies in mood disorders indicate altered numbers of neurons and glial cells. Biol Psychiatry. In press
- 129. Rajkowska G, Miguel-Hidalgo JJ, Wei J, et al. Morphometric evidence for neuronal and glial prefrontal cell pathology in major depression. Biol Psychiatry 1999;45:1085–1098
- 130. Chen RW, Chuang DM. Long term lithium treatment suppresses p53 and Bax expression but increases bcl-2 expression. J Biol Chem 1999;274: 6039–6042
- Pascual T, Gonzalez JL. A protective effect of lithium on rat behaviour altered by ibotenic acid lesions of the basal forebrain cholinergic system. Brain Res 1995;695:289–292
- 132. Arendt T, Lehmann K, Seeger G, et al. Synergistic effects of tetrahydroaminoacridine and lithium on cholinergic function after excitotoxic basal forebrain lesions in rat. Pharmacopsychiatry 1999;32: 242–247
- Nonaka S, Chuang DM. Neuroprotective effects of chronic lithium on focal cerebral ischemia in rats. Neuroreport 1998;9:2081–2084
- Chuang DM, Wei H, Qin Z, et al. Lithium inhibits striatal damage in an animal model of Huntington disease. Presented at the 241st annual meeting of the Society of Neuroscience; Oct 25, 1999; Miami Beach, Fla
- Klein PS, Melton DA. A molecular mechanism for the effect of lithium on development. Proc Natl Acad Sci U S A 1996;93:8455–8459
- Bijur GM, De Sarno P, Jope RS. Glycogen synthase kinase-3β facilitates staurosporine- and heat shock-induced apoptosis: protection by lithium. J Biol Chem 2000;275:7583–7590
- Eriksson PS, Perfilieva E, Bjork-Eriksson T, et al. Neurogenesis in the adult human hippocampus. Nat Med 1998;4:1313–1317
- Kempermann G, Gage FH. Experience-dependent regulation of adult hippocampal neurogenesis: effects of long-term stimulation and stimulus withdrawal. Hippocampus 1999;9:321–332
- Chen DF, Schneider GE, Martinou JC, et al. Bcl-2 promotes regeneration of severed axons in mammalian CNS. Nature 1997;385:434–439
- 140. Moore GJ, Bebchuk JM, Hasanat K, et al. Lithium increases N-acetylaspartate in the human brain: in vivo evidence in support of bcl-2's neurotrophic effects? Biol Psychiatry. In press
- 141. Moore GJ, Bebchuk JM, Wilds IB, et al. Pharmacologic increase in human gray matter. Lancet. In press
- Duman RS, Heninger GR, Nestler EJ. A molecular and cellular theory of depression. Arch Gen Psychiatry 1997;54:597–606